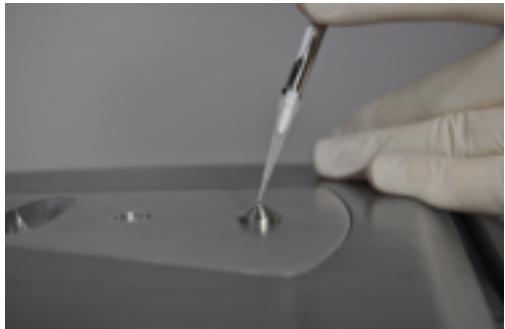


Best Practices

Technical Note 106

Introduction

Microvolume spectrophotometric measurements are now routine in many life science research labs. As with all microvolume techniques, it is important to apply best practices to ensure reliable results and reduce user error.



Sample Delivery

- Clean both sample measurement surfaces prior to making the Blank measurement.
- Use 1 μ L samples for routine measurements. Note: Select the Short Path mode for high concentration 0.5 μ L samples.
- Use a fresh aliquot for each measurement.
- Use a fresh tip to deliver each sample aliquot.
- Avoid introducing bubbles when pipetting samples onto the measurement surfaces.
- Use a dry lab wipe to remove the sample from both the top and bottom surface immediately after each measurement.
- Ensure sample concentrations fall within absorbance limits of the instrument for accurate results.

Sample Preparation

- Ensure sample isolation protocols are optimized and samples are purified prior to measurement.
- Ensure sample solutions are homogenous and well mixed before sampling.

Buffer Compatibility

- Use the same buffer a sample is suspended in for the Blank measurements.
- Avoid buffers that contain components with strong absorbances at the wavelength of interest.
- Colorimetric methods such as the Pierce 660 nm assay are recommended for proteins suspended in RIPA buffers. Contact the reagent manufacturer for buffer compatibility.

Sample Surface

- Use dH₂O, not detergents or alcohol, for routine sample measurement surface cleaning.

Cuvette Compatibility (DS-11+)

- Use a cuvette with a Z height of 8.5 mm.
- Use clean cuvettes for each sample measurement. Clean cuvettes according to the manufacturer's recommended protocol.
- Use UV transparent quartz cuvettes for nucleic acid and Protein A280 measurements.
- Ensure the cuvette is inserted in the proper orientation.

